

A High-Throughput and Low-Cost 3D Imaging System for Flowing *Escherichia Coli*

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Abstract—In this paper, we present a novel design of low-cost, high-throughput hologram-based 3D imaging system towards the development of a detection system for *Escherichia coli* (*E. coli*). This focusing-free system can image food samples flowing in a transparent microchannel under illumination of RGB lasers through a pinhole, and yields their 3D structures in real-time using hologram-based reconstruction algorithms. We solve two key technical problems in order to realize the system: 1) implementation of a high-speed (200fps) imaging system for capturing full-frame holograms of cells flowing in microchannels; 2) reconstruction of 3D structure of *E. coli* by characterizing the holograms under low-cost incoherent light sources such as the incoherent laser diodes. Market-dominating food pathogen detection systems using the polymerase chain reaction (PCR) are very slow (10-20 hours/ml) in addition to their high costs. Moreover, they cannot detect the viable but non-culturable (VBNC) *E. coli*, which orchestrated the recent massive and deadly food poisoning outbreaks in Europe and North America. The estimated cost of the proposed system is less than US\$11,000 and its speed is over 1,000 times faster than the existing methods.

I. INTRODUCTION

E. coli has been a key indicator for fecal contamination in food [1], [2]. Modern detection methods like the real-time polymerase chain reaction (PCR) [3], [4], [5] and the nucleic acid sequence based amplification (NASBA) [8,9] can detect *E. coli* but they take 10 to 20 hours for results and cost more than a million Hong Kong Dollars. Worse still, a massive food poisoning outbreak in Germany and North America revealed that, the existing *E. coli* detection methods are worthless in finding the virulent serogroups of viable-but-non-culturable (VBNC) *E. coli* [6], [7], [8], [9]. As a result, a low-cost and high-throughput detection system for *E. coli* and VBNC *E. coli* is desperately needed worldwide.

In this paper, we present our works in developing a low-cost and high-throughput 3D imaging system for detection of *E. coli*, including the hard-to-detect VBNC *E. coli*. The fundamental working principle is similar to the digital holographic microscopes (DHMs) in the market, which employ Gabor's hologram theory [10] for reconstruction of sample's 3D structure. However, our design, methodology and functionality are totally different. The proposed system leverages low-cost, incoherent, red, green and blue laser diodes as light sources, and yields the sample's 3D structure with chromatic features in real-time. In addition, the imaging component in the system comprises a high-end scientific CMOS and a field-programmable gate array (FPGA) for a stable image

acquisition and processing. These configurations and functionalities facilitate an innovative *E. coli* detection system with a cost of US\$11,000 and a detection 1,000 times faster than the existing products in the market.

Modern detection methods for *E. coli* include the immunoassay methods [11], the real-time polymerase chain reaction (real-time PCR) [3] and the nucleic acid sequence based amplification (NASBA) [4], [12]. The immunoassay methods normally take 4 to 7 days. Real-time PCR and NASBA are genetic amplification methods and now at front-line of food pathogen detection worldwide to detect a wide range of virulent microorganisms. The genetic amplification methods involve several key steps like samples enrichment, denaturation, annealing and extension. NASBA does not require a precise and hierarchical thermal control as found in the real-time PCR. Nonetheless, both methods require a long sample preparation time including the process of cell culturing. A recent development in the immunomagnetic separation (IMS) [2], [13], [14], [15] can speed up the overall detection time, but the real-time PCR and NASBA are still sensitive to inhibitors and require extensive sample preparation in addition to their extremely high cost. A cell-phone based *E. coli* detection method has recently been proposed and evaluated [16], [17], [18]. Their method relies on dyeing fluorescent particles [19], particularly the streptavidin conjugated quantum dots, along with the biotinylated anti-*E. coli* antibodies and other antibodies for capture and detection, on *E. coli* for the detection. After incubation and sample preparation, they use the camera in a cell-phone to look for the fluorescent spots in the dyed samples. The overall detection time takes 4-5 hours. The immunoassay method is recently re-visited and showed promises of fast *E. coli* detection in less than an hour [20]. However, it requires a rather high concentration of nearly 50 CFUs per milli-liter of *E. coli* in samples for detection.

In this paper, we address three key challenges that pertain to the development of the proposed system. 1) We characterize the hologram under incoherent light sources to rectify the undesired fringes that obscures the reconstruction of 3D structures of *E. coli*. 2) We develop a method to combine three holograms, that are illuminated by red, green and blue lasers respectively, then yield 3D chromatic structures of the samples in real-time. 3) We implement a high-speed (200fps) imaging system for capturing the full-frame holograms.

From a technology perspective, we adapt the hologram-based reconstruction algorithms developed in Fourier optics [21], specifically the Fresnel-Kirchhoff diffraction integral and the Rayleigh-Sommerfeld integral [22], with a

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series of image processing techniques to synthesize 3D chromatic images of the microorganism in food samples. The technology of color hologram generation is widely adapted in the fields of picture coding and printing industry [23], but it is not further developed to accommodate in the holographic microscopy or the reconstruction of hologram.

The paper is arranged as follows: Section II explains the principle of holography, and Section II-B details methods for the reconstruction of holograms. Section II-C includes the reconstruction method with objective lens. Section II-D discusses the algorithms for the generation of depth maps. Section III gives and analyzes the experimental results. Finally, Section IV closes with a conclusion.

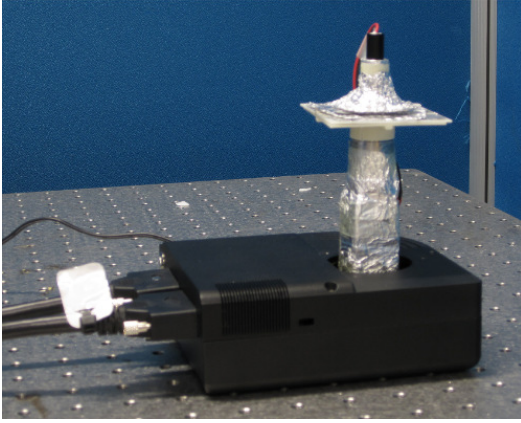


Fig. 1. A prototype of our high-throughput, low-cost 3D imaging system for the detection of *E. coli*. Only one laser diode is incorporated in this configuration. The imaging system foundation originates from Pho Imaging's Oria imaging system (#ORIA-2-CL-C-S), and is modified to fit our application.

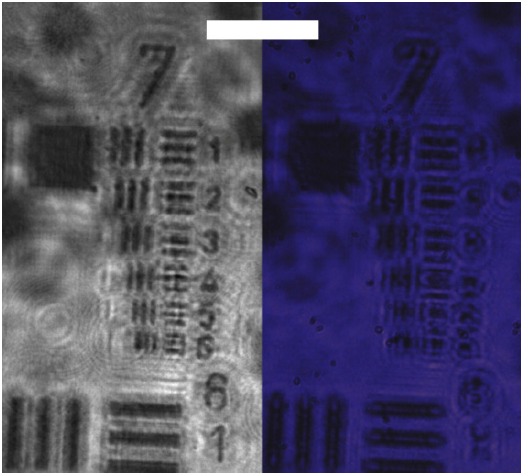


Fig. 2. (Left) A reconstructed hologram; (Right) The original hologram of a USAF test object captured by our system. The length of white bar is of 50 μ m.

II. RECONSTRUCTION ALGORITHM

A. A Brief Review of Holography

Microscope holography is first introduced by Gabor in 1948 [24], which at first was used to avoid spherical

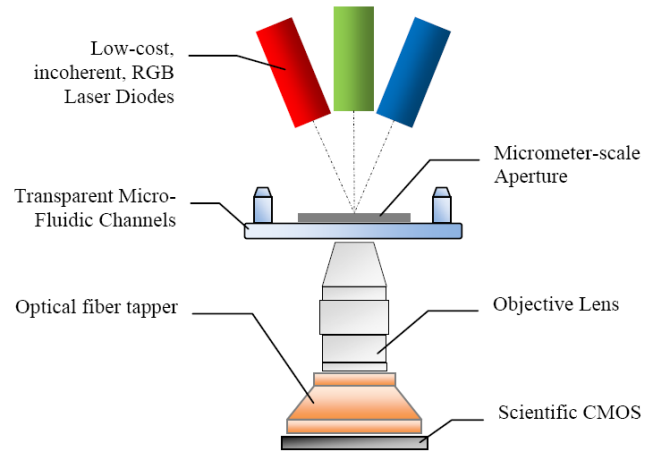


Fig. 3. Schematics of our system.

aberrations of electron lenses. The whole recording system includes a point source, an object and a photographic plane. The primary wave emitted from the point source (namely the reference wave), and the secondary wave scattered by the object (namely the object wave) interfere together at the holographic plane, and create an intensity recording - the hologram. Over the years, this holography principle received enormous attention in optical microscopy as the 3D information of the object can be, in principle, retrieved by a single image of the object's hologram.

The holography principle, however, suffered from an inherent noise coined as the twin image problem. Many aimed at eliminating this noise and in 1962, Leith and Upatnieks [25] addressed the twin image by proposing an off-axis setup to spatially separates the twin image. Other methods like Phase Shifting Holography [26], [27] are also proposed to solve this problem by altering the phase of the reference wave and reconstructing using multiple holograms.

In the late 1960s and early 1970s, the digital holography (DH) [28], [29], [30] appeared and replaced the photographic plane with a digital sensor and the reconstruction is carried out on computers. However, the computers and sensors were incompetent to perform reconstruction in real time. Under the fast-paced development of the computation capacity and the booming digital imaging resolution, DH revived in 1990s and was advanced to numerically suppress the twin image problem using various algorithms. Currently, the modern DH method is a low-cost and focus-free approach to record, reconstruct and observe the 3D information of an object.

B. Inline Digital Hologram

Figure 4 illustrates the basic setup of Gabor's inline holography [24]. In general, the numerical reconstruction method of this setup can be divided into two types. The first one is based on the Kirchoff-Helmholtz transform [31], [32], [33]. The second one is based on the Fresnel-Kirchhoff integral [34], [35], [36]. Here, we adapt the latter approach and analyze Gabor's inline holography using an operator

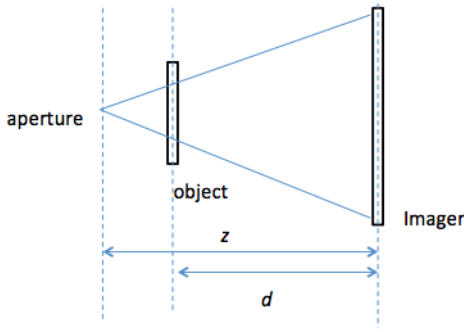


Fig. 4. Schematic of Gabor inline holography

provided by Goodman [21]. For the simplicity of this paper, we only consider 1-D case in the following analysis. The reader should note that this operator approach is based on the paraxial approximation, and omits most of the constant coefficients (e.g. The spherical wave is described as $e^{j\frac{k}{2z}x^2}$ instead of $\frac{1}{r}e^{jkr}$, where k is also the wave number, z is the distance between the point source and the plane and $r = \sqrt{x^2 + z^2}$). To demonstrate the whole process, we first briefly introduce the operators needed in this approach.

The definition of the operator Q is

$$Q[c]\{U(x)\} = e^{j\frac{k}{2}cx^2}U(x) \quad (1)$$

where $k = 2\pi/\lambda$ and c is an inverse length.

The definition of the operator V is

$$V[b]\{U(x)\} = b^{1/2}U(bx) \quad (2)$$

where b is dimensionless.

The definition of the operator R is

$$R[d]\{U(x_1)\} = \frac{1}{\sqrt{j\lambda d}} \int_{-\infty}^{\infty} U(x_2) e^{j\frac{k}{2d}(x_2 - x_1)^2} dx_2 \quad (3)$$

where d is the distance of propagation and x_2 is the coordinate that applies after propagation.

Optical lenses can be represented by the operator Q which changes the phase of the incident light. We can also convert between the plane wave and the spherical wave using operator Q . Operator V is a simple scaling operator while Operator R represents free space propagation which can be seen as an approximation of the Huygens-Fresnel principle. With these operators above we can formulate the recording process of the holograms as follows. Let P be the operator of this system, we have,

$$P = R[d]Q\left[\frac{1}{z-d}\right] \quad (4)$$

The object is illuminated by a point source that is $(z-d)$ away from the object. Then, the light propagates over the distance of d before it was imaged by a digital imager. We can rewrite (4) into (5) and (6) using the relationships between different operators as shown in [21],

$$P = Q\left[\frac{1}{z}\right]R\left[\frac{z}{z-d}d\right]V\left[\frac{z-d}{z}\right] \quad (5)$$

$$P = Q\left[\frac{1}{z}\right]V\left[\frac{z-d}{z}\right]R\left[\frac{z-d}{z}d\right] \quad (6)$$

We can design an inverse system P^{-1} based on any of the three expressions of the operators. In principle, we can fully reconstruct an object on object plane from a hologram using any of the inverse systems designed below if the amplitude and phase information are given at the imager's plane. However, an imager generally can only record the amplitude information of the light field. Without special setup like the off-axis holography, we assume that the wave field at the imager's plane is a plane wave or a spherical wave when the amplitude of the object wave is much smaller than the reference wave.

a) *Method 1 - Reconstruction using (4)*: From (4), we can reverse the hologram recording process and have,

$$P^{-1} = Q\left[-\frac{1}{z-d}\right]R[-d] \quad (7)$$

Due to the loss of the phase information, the hologram cannot be reconstructed using (7), unless we apply a Q operator at the beginning and convert the recorded hologram into a light field of ideal spherical wave. Therefore, (7) is rewritten to a more practical representation,

$$P^{-1} = Q\left[-\frac{1}{z-d}\right]R[-d]Q\left[\frac{1}{z}\right] \quad (8)$$

b) *Method 2 - Reconstruction using (5)*: If we denote the transmittance of the object as $O(x)$, we can derive the light field on the imager's plane by applying the system operator P . Then the recorded hologram can be described by, (9).

$$\begin{aligned} H(x) &= |P\{O(x)\}| \\ &= \left|Q\left[\frac{1}{z}\right]R\left[\frac{z}{z-d}d\right]V\left[\frac{z-d}{z}\right]\{O(x)\}\right| \\ &= \left|R\left[\frac{z}{z-d}d\right]V\left[\frac{z-d}{z}\right]\{O(x)\}\right| \end{aligned} \quad (9)$$

The light is propagating as a plane wave, so we can directly backpropagate over the distance of $\frac{z-d}{z}d$ and a magnified object image as shown in (10) can be obtained,

$$\begin{aligned} R\left[-\frac{z}{z-d}d\right]\{H(x)\} &\approx R\left[-\frac{z}{z-d}d\right]R\left[\frac{z}{z-d}d\right]V\left[\frac{z-d}{z}\right]\{O(x)\} \\ &= V\left[\frac{z-d}{z}\right]\{O(x)\} \end{aligned} \quad (10)$$

Therefore, we can have

$$P^{-1} = R\left[-\frac{z}{z-d}d\right] \quad (11)$$

c) *Method 3 - Reconstruction using (6)*: The reconstruction steps based on (6) is similar to the Method 2 except that before performing the back propagation, the holograms must be virtually magnified by setting the pixel size of the imager $\frac{z-d}{z}$ times smaller than its original dimension.

Comparing the three reconstruction methods above, the Method 2 requires the least computation effort and generates a magnified object image which can help in inspecting small objects like the E. coli. Therefore we adapt this approach to reconstruct the 2-D image of the hologram.

C. 2-D Reconstruction with Objective Lens

To further amplify the object, our system is equipped with an objective lens and is placed between the object and the imager. Figure (5) illustrates the design of this setup. The

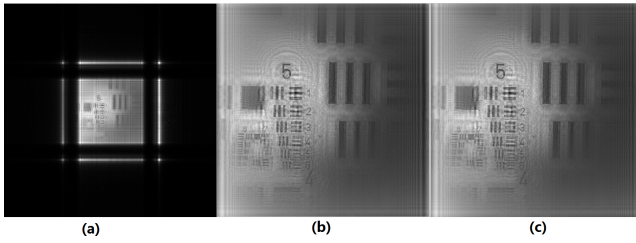


Fig. 7. Reconstruction results using Gabor's inline holography (a) By (4); (b) By (5); (c) By (6)

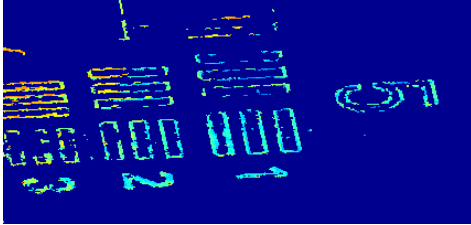


Fig. 8. Results of depth map generation

method yields a reconstructed image of the testing object using the original image size while the other two methods yield magnified images. Figure 2 exhibits the results by our system with a 40x oil immersion objective lens. We can observe the Group 6 and 7 which is the smallest group of our testing target. The shortest length width among these two groups is about 2.19 μ m. The computation time is less than 20ms for the reconstruction of a single, full-size hologram. The results of the depth map is given in Fig. 8. The resolution can be improved by replacing the objective lens with a higher magnification one. The results demonstrate that the proposed system can image micro-object with a resolution of about 2 μ m and is expected to achieve a resolution of 0.8 μ m when using a 100x objective lens. As the size of E. coli is around 1 to 3 μ m, the system is capable of capturing of these samples with distinguishable resolution. We will further develop the system, particularly on the processing and classification of imaged samples using their chromatic and geometric features in complex food samples.

IV. CONCLUSION

The viable but non-culturable (VBNC) E. coli is fatal to human and is undetectable using the existing detection systems in food industry. The proposed focusing-free 3D imaging system can detect micro-object as small as 1 to 2 μ m with a throughput of 40 minutes per ml, and hence can be further developed then deployed to detect the virulent strains of pathogenic E. coli at a speed that can be 1,000 times faster than today's top-notched E. coli detection products. At the core of our system is a high-speed, large field-of-view imaging system and a novel design of chromatic laser illumination setup. By leveraging holographic principles, we

adapt Gabor's algorithm to yield 3D structures of the flowing E. coli in a micro-channel, and lay a promising foundation for the development of high-throughput, low-cost detection systems for E. coli in food samples.

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